



A QSAR Study of New Caffeine Derivatives with Epithelial Anticancer Activity

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Authors' contributions

This investigation was performed in collaboration with all authors. Authors LKSG, JBV, FSB and CBRS designed the study, wrote the protocol, involved in writing the first draft, participated in data collection. Authors NSRS, CFS, JSC and WJCM managed the literature search, analyses of the study and manuscript preparation. Authors CHTPS, LISH and CBRS performed data interpretation and were actively involved in reading the manuscript. All authors read and approved the final manuscript.

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ABSTRACT

Aims: To study and propose new caffeine derivatives with epithelial anticancer activity using quantum chemistry methods and multivariate analysis (PCA, HCA, PLS and PCR).

Place and Duration of Study: Laboratory of Modeling and Computational Chemistry at Federal University of Amapá (UNIFAP), Macapá, Brazil, between March 2014 and February 2015.

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Methodology: Caffeine and 31 derivatives with epithelial anticancer activity were selected from the literature, and modeled with the GaussView 3.0 program. The optimization was performed using the DFT method and B3LYP/6-31G** base set implemented in the Gaussian 03 program. Principal component analysis (PCA) and hierarchical cluster analysis (HCA) were employed to select the molecular descriptors related to epithelial anticancer activity. The Pearson correlation between activity and important descriptors were used for the regression partial least squares (PLS) and principal component regression (PCR) models built, and these models were used to predict the anticancer activity of fourteen new caffeine derivatives (test set) with unknown activity.

Results: The PCA results showed that the descriptors related to the compounds with anticancer activity were: area (A^2), distance radical 1 (dR1), distance radical 3 (dR3), radical partition coefficient 1 (logPR1) and radical partition coefficient 3 (logPR3). HCA showed similar results obtained with PCA, and the compounds were grouped in accordance with their biological activities. The results obtained with the PLS and PCR models were close, with variation between the models of $R^2=\pm 0.005$, $R^2_{\text{ajust}}=\pm 0.1998$, $s=\pm 0.0053$, $F_{(5,27)}=\pm 49.2261$, $Q^2=\pm 0.012$, $SEV=\pm 0.0117$, $PRESS=\pm 0.0473$ and $S_{PRESS}=\pm 0.0087$. The PLS model showed that eight compounds of the test set (37 and 40-46) are predicted to be more active, and they had values of $ICT_{50}<0.48\text{mM}$. However, the PCR model only seven compounds of all test sets (33-36, 38, 39 and 42) were predicted as most active, which showed values of $ICT_{50}\geq 0.48\text{mM}$, a total of 7 compounds proposed as less active of fourteen suggested compounds. However, compounds 37, 40, 41, 43, 44, 45 and 46 were the ones that had values of $ICT_{50}<0.48\text{mM}$ in both PLS and PCR models, suggesting that these new compounds in the two models are more potent than caffeine may be tested for epithelial anticancer activity.

Conclusion: The PLS and PCR models showed good predictive ability. The test set showed for seven new caffeine compounds satisfactory results for epithelial anticancer activity. This strategy is fundamental for use in experimental syntheses and biological evaluation, and to understand the structural requirements for designing new ligands as anticancer agents.

Keywords: Caffeine; epithelial cancer; molecular modeling; B3LYP/6-31G**; QSAR.

1. INTRODUCTION

During the history man observed that plants used for therapeutic purposes. The choice of these plants for pharmaceutical applications were based more on empirical data than the scientific. Current events instigated a search for compounds that stand out from studies, by means of modifications to these structures, promoting better activity and reduced undesirable effects such as toxicity [1].

Caffeine (1,3,7-trimethylxanthine), see Fig. 1, is a chemical compound classified as alkaloid (basic character substance derived from plants containing in its formula, basically nitrogen, oxygen, hydrogen and carbon) pertaining to the group xanthine (organic substance, total nitrogenous, existing in the cardiac muscle, urine, and in various organs in some plants) [2]. Caffeine is found in some plants and used for consumption in beverages, as an infusion, as a stimulant, extremely soluble in hot water, and has no smell and has bitter taste [3].

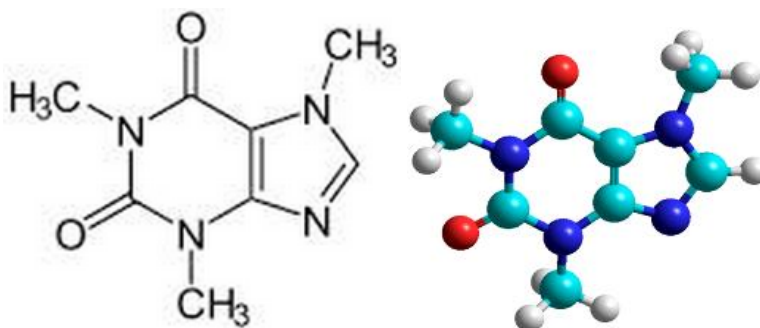


Fig. 1. Structural formula of caffeine (1,3,7-trimethylxanthine) in 2D and 3D format

Currently a variety of factors have motivated the search for new drugs of vegetable origin, among these stands out the discovery of drugs to combat cancer successfully [4]. Chaturvedi [5] relates that presently the antitumor action is the most extensively studied biological activity of plants, where studies have shown that they are able to combat tumors by selective alkylation, controlling and inhibiting cell division. These factors and cellular functions lead the cells to die by apoptosis. Cancer is called malignant neoplasm or malignant tumor, where the origin are due to genetic alterations can be oncogene activation, tumor suppressor genes inactivation, inactivation of genes responsible by apoptosis, mutations produced by chemical, physical and biological agents, and characterized by loss of function from the absence of differentiation, uncontrolled proliferation, invasiveness of adjacent tissues and metastasis [6-8].

On a worldwide there was an increase to 14.1 million new cases of different types of cancer in 2012, these 8.2 million deaths caused [9]. The types most frequently diagnosed were lung (13.0% of the total), breast (11.9%), and colon and rectum (9.7%). The most common determinants of death were lung (19.4% of total), liver (9.1%) and stomach (8.8%) cancers [10]. Through the need to eradicate this type of pathology, caffeine shows meaningful data that can come to combat the epithelial cancer. This pathology occurs in countries that possess favorable climatic conditions, where excessive sun exposure is the main risk factor for epithelial cancer. Among the countries with tropical climates stand out Brazil, followed by Australia [11,12].

Epithelial cancer better known as skin cancer is characterized by the uncontrolled growth of a group of cells of an organism, and this pathology is the result mainly of genetic alterations, environmental factors and lifestyle. This type of cancer is very incident in the world and the entire population is susceptible [13]. Epithelial tissue has several different layers and depending on where occurs at the cellular disorder cancer receives several different nomenclatures, such as: basocellular carcinoma, squamous cell carcinoma and malignant melanoma [14].

Ultraviolet radiation (UVR) is a complete carcinogen, and initiates the malignization process by mutations in the DNA and promotes the development of cancer by inflammatory process inherent to cumulative UVR exposure

[15]. The UV rays contribute to the development of both forms of skin cancer: melanoma and non-melanoma. The non-melanocytic tumors are associated with cumulative sun damage, while melanocytic are intimately associated with intense episodes of excessive sun exposure, resulting in sunburn [16]. It is observed that skin manifestations present an evolutionary spectrum of appearance, in this order: burning, skin thickening, hyperchromic spots, fine lines, deep wrinkles and actinic [15].

The caffeine has stimulating activity on the central nervous system, cardiac muscle, and gastric acid secretion. Its activity is relatively beneficial when used in moderate doses, but in excess doses used, stimulates improperly the central nervous system, causing often accented frame insomnia and irritability [17,18]. For these reasons develop new studies for the use of caffeine. As well as there are new activities of other drugs on the market, such as thalidomide.

Thalidomide was first used in 1956 in Germany, through the therapeutic activity as a sedative [19]. Over the years she was gaining the market in 40 countries include Canada and the UK, but in the United States of America (USA) was not successful, the Food and Drug Administration (FDA) did not approve their entry for security criteria not many enlightened on drugs [20,21]. Due to the discovery that the drug has teratogenic characteristics in 1961, it was totally suspended from the world market [22,23]. Although the global scandal of thalidomide about their teratogenic activity, today it is being introduced in healing of erythema nodosum leprosum (ENL), and another very interesting activity to be mentioned is combat the multiple myeloma [24]. The study presented analogs that were effective against multiple myeloma, containing less side effects of drugs, and future research will be developed with nano basis for production of less toxic drugs that can combat cancer cells [25,26].

In this paper, a QSAR study of caffeine and 31 derivatives with epithelial anticancer activity (see Fig. 2) was performed. The structures were modeled and various different molecular descriptors were calculated with B3LYP/6-31G** method. The Principal Component Analysis (PCA) and Hierarchical Cluster Analysis (HCA) were employed to select the molecular descriptors that are most related with biological activity investigated. A QSAR model was built with the Partial Least Square (PLS) and Principal

Component Regression (PCR) methods to carry out predictions of fourteen new caffeine compounds (test set) with unknown anticancer activity [27-30]. These predictions help in the interactions between molecules and their molecular targets, and to aid in future studies searching for other new anticancer drugs.

2. METHODOLOGY

The study was performed in Laboratory of Modeling and Computational Chemistry at Federal University of Amapá, Macapá, Brazil, between March 2014 and March 2015.

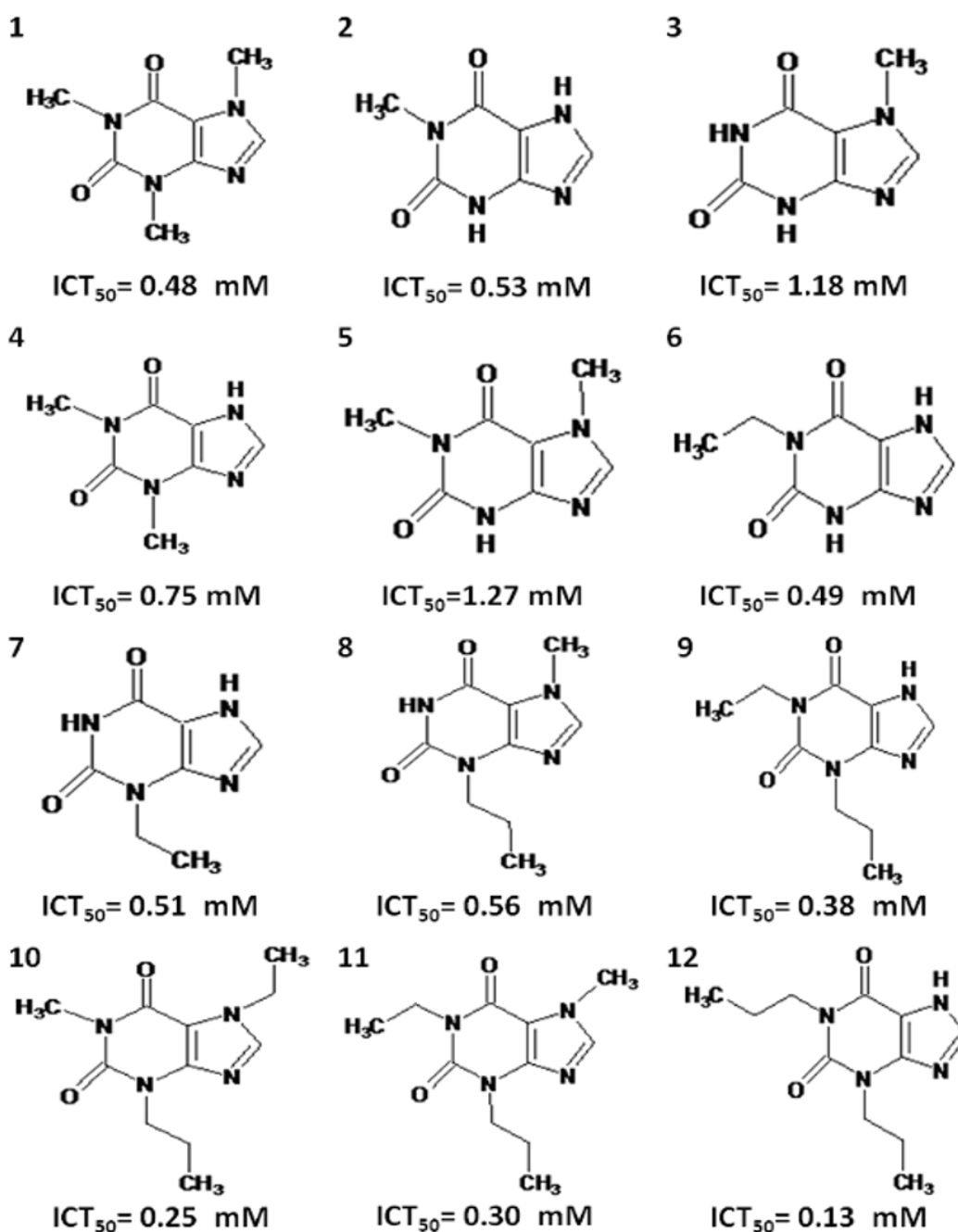


Fig. 2. Continuation

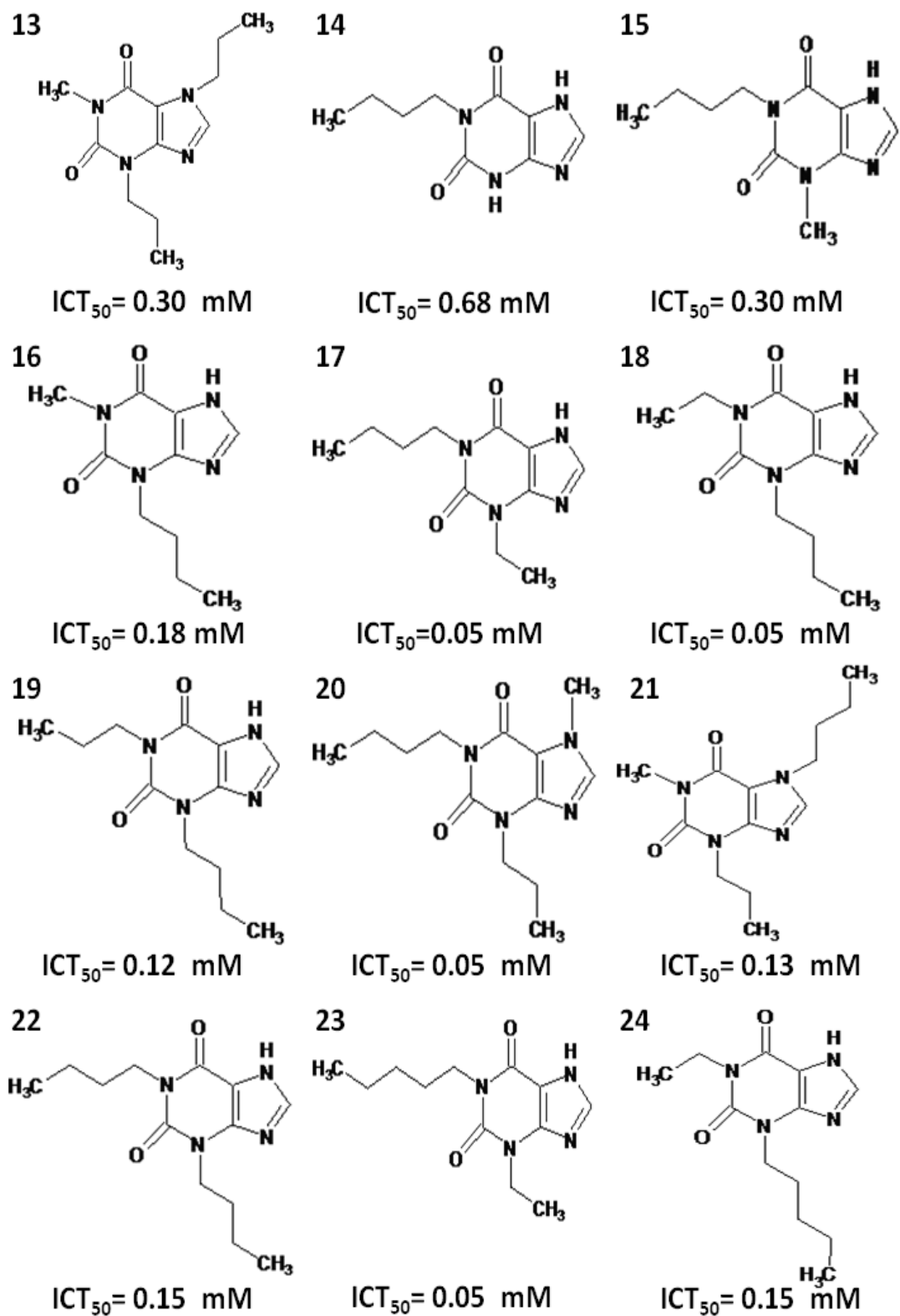


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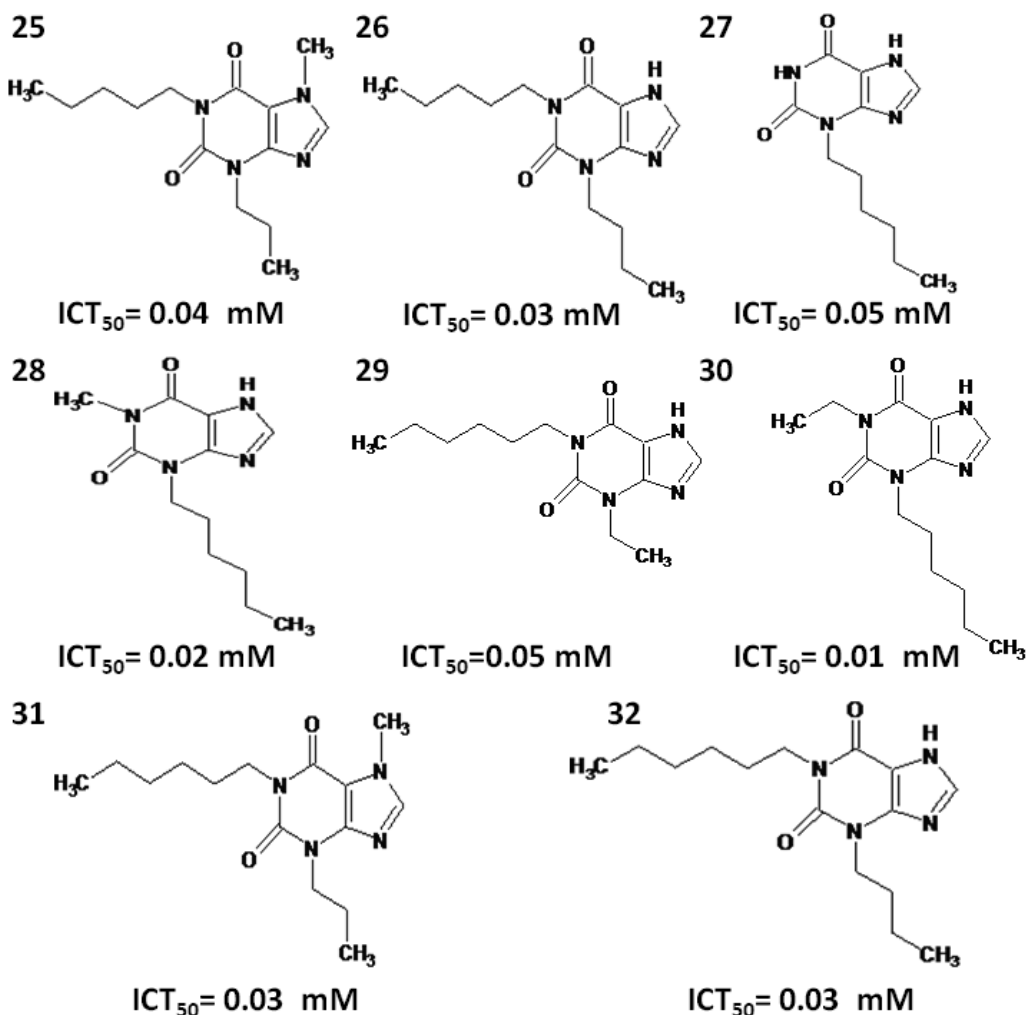


Fig. 2. Structure and biological activity of caffeine and its derivatives with epithelial anticancer activity

2.1 Compounds Studied with Epithelial Anticancer Activity

Initially, caffeine and 31 derivatives with epithelial anticancer activity were selected from the literature; see Fig. 2 [31]. The compounds were based on chemical structures, which differ in the length of the side chain alkyls (radicals). This size difference of the radicals, radical 1 (R1), radical 3 (R3) and radical 7 (R7), showed that the antitumor activity of these compounds suggested that inhibition of cell transformation to 1,3,7-trialkylxanthines depends on the number of carbon atoms for the alkyl group R1, R3 or R7. However, this study of Rogozin depicted that some xanthine analogues exert anticancer

activity in epidermal cell cultures in animal models. Therefore, it is based on prevention of epidermal growth factor (EGF). All the compounds studied have been associated with *in vivo* activity against inducing malignant transformation in epidermal cells of rats JB6 susceptible to the development (P+) C141 (JB6 P+), based on the values of 50% inhibition of cell transformation (ICT_{50}) [31].

In this paper, the compounds were classified based on the anticancer responses adopted: compounds with $ICT_{50} < 0.48$ mM, ranging from 0.0100 to 0.3800 mM, were assumed to be more potent compounds (9-13 and 15-32), and those with $ICT_{50} \geq 0.48$ mM, ranging from 0.4800 to

1.2700 mM, were considered to be less potent compounds (1-8 and 14).

2.2 Molecular Modeling of Caffeine and Its Derivatives and Calculations of Descriptors or Molecular Properties

Compounds were modeled with GaussView 3.0 program [32], following the described strategy: initially the structure of caffeine was built and optimized with the DFT method and B3LYP/6-31G** base set implemented in Gaussian 03 program [33]. After obtaining of more stable geometry of caffeine, other compounds were built, and subsequently, obtained their most stable structures with lower energy in the same method and base set of caffeine.

The molecular descriptors are important for description of molecular structure and to finding appropriate predictive models [34]. The calculations of the molecular descriptors were performed employing the following software: Gaussian 03 program, Molekel [35] and HyperChem 6.02 [36].

In our study we calculated the chemical descriptors: total energy (TE), lowest unoccupied molecular orbital energy (LUMO), a level above the energy of the lowest unoccupied molecular orbital (LUMO+1), energy of the highest occupied molecular orbital (HOMO), a level below the energy of the highest occupied molecular orbital (HOMO-1), difference in energy between HOMO and LUMO (GAP=HOMO-LUMO), Mulliken electronegativity (χ), molecular softness ($1/\eta$), molecular hardness (η), torsion angle, dihedral angle, dipole moment (μ) and partial atomic charge. The atomic charges used in this study were obtained with the key word POP=CHELPG using the electrostatic potential [37], and atomic charges offer the general advantage of being physically more satisfactory than Mulliken charges [38]. In addition, the binding orders were calculated, distance radical 1 (dR1), distance radical 3 (dR3), distance radical 7 (dR7), hydrophobic descriptors such as the partition coefficient (logP), partition coefficient for radical 1 (logPR1), partition coefficient for radical 3 (logPR3), partition coefficient for radical 7 (logPR7) of all compounds studied. The molar refractivity, polarizability molar (MP), molar volume (MV), molar area, molar mass and hydration energy (HE) descriptors were calculated with the HyperChem 6.02 program.

2.3 Variable Selection and Construction of PLS and PCR Models

After determining all descriptors, a data matrix was built to start the multivariate analysis step. Multivariate analysis step was necessary to make the standardization or autoscale of the data matrix $X=(n, m)$, formed of thirty-two (32) lines (anticancer compounds) and seventy-six (76) columns (descriptors or molecular properties for each molecule), where n is the number of compounds and m is the number of variables.

The aim of standardized data matrix is to give equal weight to each variable in mathematical terms [39]. Variable selections were based on the analysis of the Pearson correlation matrix between variables and the epithelial anticancer activity (ICT₅₀). The descriptors or molecular properties with small or no correlation were discarded. After this analysis, two complementary methods were employed (PCA and HCA) to select the properties that contribute for classification of the compounds into two groups (more potent compounds and less potent compounds) [40-42].

PCA was used to reduce the dimensionality and find descriptors that could be useful in characterizing the behavior of the compounds with epithelial anticancer activity, and analyze the natural grouping of data and discrepant samples. During the execution of PCA, several attempts were made to get a good classification. The score plot gives information about the compounds (similarity and differences), and the loading plot gives information about the variables [29,30,42-46]. The descriptors selected by PCA were used to carry out HCA and for build of PLS and PCR models.

The aim of HCA is to show the compounds distributed in similarity, and the results should confirm of the PCA. The compounds become an agglomeration type, because each compound was first defined as its own cluster, and then others were grouped together to form new clusters until all the compounds were part of a single cluster [29,30,42-46].

The QSAR models were constructed by the PCR and PLS methods based on the autoscaled data and the leave-one-out cross validation procedure [29,30,42-46]. The statistical parameters used to evaluate the quality of the models were: Prediction Residual Error Sum of Squares (PRESS), Equation (1), Standard Error of

Validation (SEV), Equation (2), total variance explained, R^2 (correlation between the estimated values predicted by the model built with the full data set and actual values of y), Q^2 (the cross-validated correlation coefficient) and S_{PRESS} (standard deviation of cross-validation) given by Equations (3)–(5), respectively [29,30,42-46].

$$PRESS = \sum_{i=1}^n (y_i - \hat{y}_i)^2 \quad (1)$$

$$SEV = \sqrt{\frac{PRESS}{n}} \quad (2)$$

$$R^2 = 1 - \left[\frac{PRESS_{cal}}{\sum_{i=1}^n (y_i - \bar{y})^2} \right] \quad (3)$$

$$Q^2 = 1 - \left[\frac{PRESS_{val}}{\sum_{i=1}^n (y_i - \bar{y})^2} \right] \quad (4)$$

$$S_{PRESS} = \frac{\sqrt{PRESS}}{n - k - 1} \quad (5)$$

In Equations (1) and (2), n is the number of compounds used for the validation model, y_i is the experimental data of the molecular properties for the sample and \hat{y}_i is the value predicted by a calibration or validation model. In Equations (3) and (4), $PRESS_{cal}$ is the Calibration Prediction Error Sum of Squares and $PRESS_{val}$ is the Validation Prediction Error Sum of Squares. Both $PRESS_{cal}$ and $PRESS_{val}$ are evaluated from Equation (1) by changing \hat{y}_i for a calibration or validation model. The values of explained variance (R^2 ajust, i.e., adjusted R^2), standard deviation (s) and F (Fisher test) were determined. The aim of the PLS and PCR methods were the constructions of a mathematical model that can be used to predict epithelial anticancer activity of fourteen new caffeine compounds (test set) with unknown anticancer activity. PCA, HCA, PLS and PCR were performed using the Pirouette 3.01 [47] and Statistica 6.2 [48] programs.

3. RESULTS AND DISCUSSION

3.1 Principal Component Analysis (PCA) Results

PCA results showed that the important molecular properties were the following: area (A^2), distance radical 1 (dR1), distance radical 3 (dR3), radical partition coefficient 1 (logPR1) and radical partition coefficient 3 (logPR3). They were selected from the data set of 76 molecular properties, and other variables were not selected because had a poor linear correlation with epithelial anticancer activity.

The values of the important molecular properties of each selected compound identified via PCA as well as the values of ICT_{50} (50% inhibition of cell transformation), are shown in Table 1. Table 1 shows the Pearson correlation matrix between the molecular properties and ICT_{50} , and the correlation between pairs of molecular properties is less than 0.9833, while the correlation between the molecular properties and ICT_{50} is more than -0.7885. The molecular properties selected by PCA represent the characteristics to separate between the more and less active with epithelial anticancer activity of the compounds.

The results of the PCA model are presented in Table 2. The model was built with three main components (3 PCs). The first principal component (PC1) describes 92.1316% of the total information, the second principal component (PC2) describes 58.8150%, and the third (PC3) 2.5021%. PC1 contains 59.4397% of the original data, and the combination of the first two components (PC1+PC2) contains 97.3850%, and all three (PC1+PC2+PC3) explain 98.9992% of the total information, losing only 1.008 of the original information. The dR3 (0.4261), logPR3 (0.4396) and A^2 (0.5642) descriptors contribute to PC1, while in PC2, the main contributors are the dR1(0.5190) and logPR1 (0.5283) descriptors.

The main components can be written as a linear combination of the selected descriptors. Mathematical expressions for PC1 (6) and PC2 (7) are shown below:

$$PC1 = 0.3963dR1 + 0.4261dR3 + 0.3869logPR1 + 0.4396logPR3 + 0.5642A^2 \quad (6)$$

$$PC2 = 0.5190dR1 - 0.4866dR3 + 0.5283logPR1 - 0.4632logPR3 + 0.0015A^2 \quad (7)$$

Table 1. Molecular properties selected by principal component analysis, experimental ICT₅₀ (mM) values and the Pearson correlation matrix

| Compounds | dR1 | dR3 | logPR1 | logPR3 | A ² | ICT ₅₀ (mM) |
|----------------|--------|--------|--------|--------|----------------|------------------------|
| 1- | 2.1060 | 2.1040 | 1.0900 | 1.0900 | 359.0799 | 0.4800 |
| 2- | 2.1090 | 0.0000 | 1.0900 | 0.0000 | 237.9100 | 0.5300 |
| 3- | 1.0140 | 1.0110 | 0.0000 | 0.0000 | 312.1600 | 1.1800 |
| 4- | 2.1080 | 2.1060 | 1.0900 | 1.0900 | 337.6099 | 0.7500 |
| 5- | 2.1080 | 1.0110 | 1.0900 | 0.0000 | 337.4100 | 1.2700 |
| 6- | 3.4390 | 0.0000 | 1.3000 | 0.0000 | 324.3999 | 0.4900 |
| 7- | 1.0140 | 3.4340 | 0.0000 | 1.3000 | 333.1000 | 0.5100 |
| 8- | 1.0140 | 4.6730 | 0.0000 | 1.6900 | 386.5899 | 0.5600 |
| 9+ | 3.4380 | 4.6730 | 1.3000 | 1.6900 | 411.2399 | 0.3800 |
| 10+ | 2.1080 | 4.6730 | 1.0900 | 1.6900 | 436.8900 | 0.2500 |
| 11+ | 3.4380 | 4.6740 | 1.3000 | 1.6900 | 435.5799 | 0.3000 |
| 12+ | 4.6770 | 4.6740 | 1.6900 | 1.6900 | 449.4800 | 0.1300 |
| 13+ | 2.1080 | 4.6740 | 1.0900 | 1.6900 | 468.4800 | 0.3000 |
| 14- | 5.9860 | 0.0000 | 2.0900 | 0.0000 | 388.3399 | 0.6800 |
| 15+ | 5.9850 | 2.1050 | 2.0900 | 1.0900 | 422.6600 | 0.3000 |
| 16+ | 2.1080 | 5.9820 | 1.0900 | 2.0900 | 422.6400 | 0.1800 |
| 17+ | 5.9850 | 3.4350 | 2.0900 | 1.3000 | 445.6099 | 0.0500 |
| 18+ | 3.4380 | 5.9820 | 1.3000 | 2.0900 | 446.0599 | 0.0500 |
| 19+ | 4.6760 | 5.9810 | 1.6900 | 2.0900 | 477.6000 | 0.1200 |
| 20+ | 5.9860 | 4.6740 | 2.0900 | 1.6900 | 497.5499 | 0.0500 |
| 21+ | 2.1080 | 4.6730 | 1.0900 | 1.6900 | 501.8099 | 0.1300 |
| 22+ | 5.9840 | 5.9810 | 2.0900 | 2.0900 | 508.4899 | 0.1500 |
| 23+ | 7.2450 | 3.4340 | 2.4900 | 1.3000 | 476.7999 | 0.0500 |
| 24+ | 3.4380 | 7.2420 | 1.3000 | 2.4900 | 476.6600 | 0.1500 |
| 25+ | 7.2460 | 4.6740 | 2.4900 | 1.6900 | 533.3400 | 0.0400 |
| 26+ | 7.2450 | 5.9820 | 2.4900 | 2.0900 | 541.2100 | 0.0300 |
| 27+ | 1.0140 | 8.5410 | 0.0000 | 2.8800 | 459.5100 | 0.0500 |
| 28+ | 2.1090 | 8.5420 | 1.0900 | 2.8800 | 484.2000 | 0.0200 |
| 29+ | 8.5460 | 3.4350 | 2.8800 | 1.3000 | 510.7399 | 0.0500 |
| 30+ | 3.4380 | 8.5410 | 1.3000 | 2.8800 | 511.2000 | 0.0100 |
| 31+ | 8.5460 | 4.6740 | 2.8800 | 1.6900 | 564.5700 | 0.0300 |
| 32+ | 8.5440 | 5.9830 | 2.8800 | 2.0900 | 573.6500 | 0.0300 |
| dR1 | | 0.0283 | 0.9635 | 0.0660 | 0.6515 | -0.4976 |
| dR3 | | | 0.0056 | 0.9833 | 0.6988 | -0.6986 |
| logPR1 | | | | 0.0523 | 0.6302 | -0.4984 |
| logPR3 | | | | | 0.7114 | -0.7574 |
| A ² | | | | | | -0.7885 |

Fig. 3 shows the scores for the 32 compounds studied. According to the graph, PC1 separates the compounds between more and less potent. The most potent compounds are shown in the right part (9-13 and 15-32), while the less potent compounds are in the left side of the graph (1-8 and 14).

Fig. 4 shows the loadings for the five (5) descriptors that are important for the classification of compounds. The most potent compounds have high contributions of the A² and logPR3 descriptors, while less potent compounds

has a high contribution of the dR1, dR3 and logPR1 descriptors. Thus, the dR1, dR3 and logPR1 descriptors are responsible for the location of less potent compounds at the left side of the graph. The A² descriptor represents the more potent compounds in the right part of the graph. Fig. 3 also shows that the higher the contribution of the A² and logPR3 descriptors in the first principal component, i.e., the higher value for a certain compound, the higher score value will be, indicating that the compound is more potent than others.

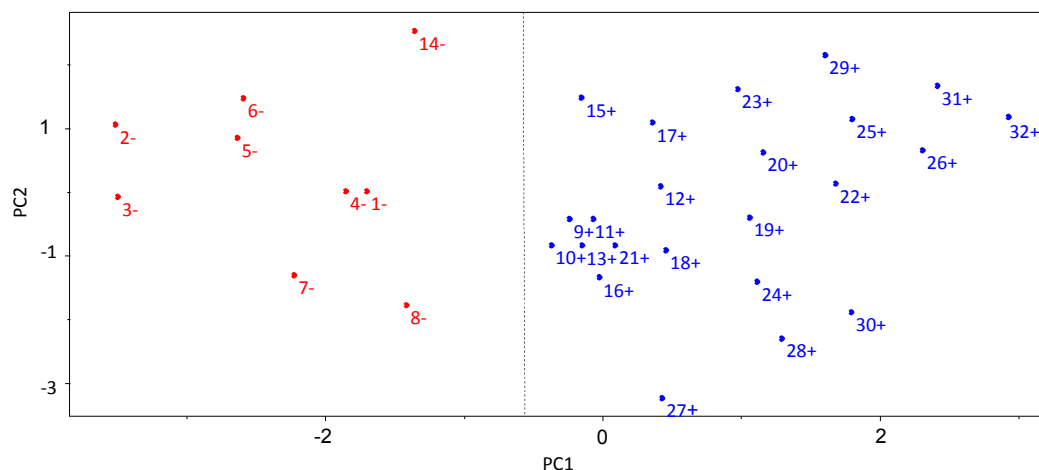


Fig. 3. Plot of PC1–PC2 scores for caffeine and derivatives with epithelial anticancer activity. Positive values indicate more potent analogs (in blue) and negative values indicate less potent analogs (in red)

3.2 Hierarchical Cluster Analysis (HCA) Results

The statistical analysis used in this study should group similar compounds into categories. The categories are represented by a two-dimensional diagram known as dendrogram which illustrates the fusions or division made in each successive analysis phase. Samples (compounds) are represented by the branches in the lower part of the dendrogram. The similarity between the groups is given by the length of its branches, thus compounds with low similarity have long branches, whereas compounds of high similarity have short branches. HCA method classified the compounds in two classes (more active and less active), and was based on the Euclidean distance and the complete method [49,50]. The descriptors used to carry out HCA were the same as for the PCA method, that is, area (A^2), distance radical 1 (dR1), distance radical 3 (dR3), radical partition coefficient 1 (logPR1) and radical partition coefficient 3 (logPR3).

The dendrogram in Fig. 5 shows the HCA result as well as the compounds separated into two classes. The scale of similarity varies from 0 for samples with no similarity and 1 for samples with identical similarity. When analyzing the dendrogram, some conclusions can be made, although the compounds show some structural diversity.

HCA showed similar results obtained by PCA. Compounds were grouped according to their biological activities. The most potent compounds

are 9, 10, 11, 12, 13, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31 and 32. The less potent compounds are 1, 2, 3, 4, 5, 7, 8, e 14. Compound 30 has the lowest value of $ICT_{50}=0.0100$, among the compounds classified as most potent. Whereas, the compound 5 has the highest value of $ICT_{50}= 1.2700$, while the variation between the activities of the compounds 5 and 30 is ± 1.2600 between them.

3.3 Partial Least Squares (PLS) and Principal Component Regression (PCR) Results

The statistical quality [49] of the PLS and PCR models was calibrated by parameters: squared correlation coefficient (R^2), explained variance (R^2 ajust), standard deviation (s), variance ratio (F), cross-validated correlation coefficient (Q^2), standard error of validation (SEV), predicted residual error sum of squares (PRESS) and standard deviation of cross-validation (S_{PRESS}) [50–52]. The best regression models were selected based on high values of R^2 , R^2 ajust, Q^2 and F (a statistic of assessing the overall significance) and low values of s, SEV, PRESS and S_{press} .

The calculated properties and the experimental values for the compounds were used to build the PLS and PCR models (see Table 3). The models built were based on three latent variables and 32 compounds. The equations obtained for PLS (Equation (8)) and PCR (Equation (9)) models are the following:

$$\text{ICT}_{50} = -0.1685\text{dR1} - 0.2366\text{dR3} - 0.1688\log\text{PR1} - 0.2564\log\text{PR3} - 0.2670\text{A}^2 \quad (8)$$

$n = 32$, $R^2 = 0.9260$, $R^2_{\text{ajust}} = 0.9157$, $s = 0.1587$, $F_{(5,27)} = 65.0702$, $Q^2 = 0.8831$, $\text{SEV} = 0.1867$, $\text{PRESS} = 0.7056$, $S_{\text{PRESS}} = 0.0191$.

$$\text{ICT}_{50} = -0.1955\text{dR1} - 0.2102\text{dR3} - 0.1908\log\text{PR1} - 0.21681\log\text{PR3} - 0.2783\text{A}^2 \quad (9)$$

$n = 32$, $R^2 = 0.9210$, $R^2_{\text{ajust}} = 0.7159$, $s = 0.1640$, $F_{(5,27)} = 15.8441$, $Q^2 = 0.8951$, $\text{SEV} = 0.1750$, $\text{PRESS} = 0.7529$, $S_{\text{PRESS}} = 0.0218$.

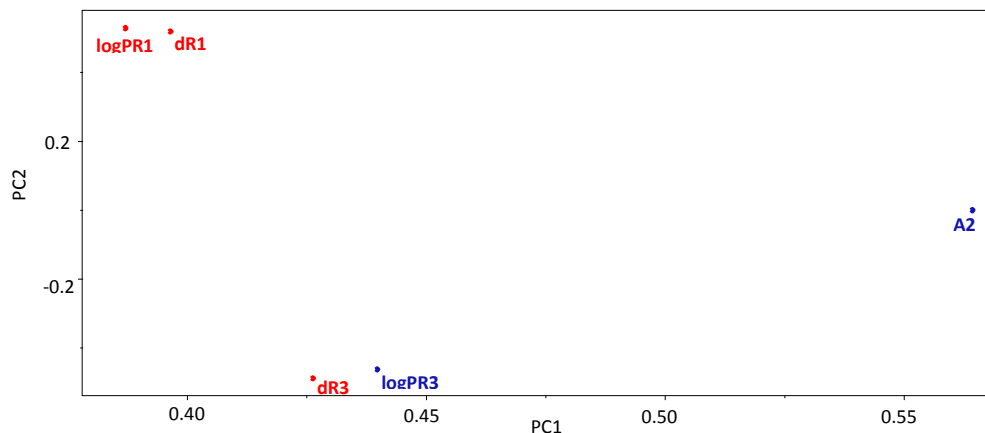


Fig. 4. Plot of the PC1–PC2 loadings with the five descriptors selected to build the PLS and PCR models of caffeine and derivatives with biological activity against epithelial cancer

Table 2. Principal component analysis of the SAR model and contribution of selected descriptors based on step multivariate analysis

| Parameters | Main component | | |
|-------------------------|----------------|---------|---------|
| | PC1 | PC2 | PC3 |
| Variance (%) | 92.1316 | 58.8150 | 2.5021 |
| Cumulative variance (%) | 59.4397 | 97.3850 | 98.9992 |
| Molecular descriptors | Contribution | | |
| | PC1 | PC2 | PC3 |
| dR1 | 0.3963 | 0.5190 | |
| dR3 | 0.4261 | -0.4866 | |
| logPR1 | 0.3869 | 0.5283 | |
| logPR3 | 0.4396 | -0.4632 | |
| A ² | 0.5642 | 0.0015 | |

The results obtained with the PLS and PCR models were close, with variation between PLS and PCR of $R^2 = \pm 0.005$, $R^2_{\text{ajust}} = \pm 0.1998$, $s = \pm 0.0053$, $F_{(5,27)} = \pm 49.2261$, $Q^2 = \pm 0.012$, $\text{SEV} = \pm 0.0117$, $\text{PRESS} = \pm 0.0473$ and $S_{\text{PRESS}} = \pm 0.0087$. The quality of the models was demonstrated by comparing the measured and the predicted activities. The validation errors obtained by the leave-one-out cross-validation method are shown in Table 3. For the PLS model, only four compounds (2, 3, 5, 17 and 32) had high validation errors, and the PCR model

yielded five compounds (2, 3, 5, 27 and 32) with high residual values.

The measured versus predicted values using our PLS and PCR models are presented in Figs. 6a,b, respectively. The PLS and PCR plots identify compounds with higher activity (blue) and compounds with lower activity (red). The validation parameters support the fact that the models are efficient and hence satisfactory given the complexity of the anticancer mechanisms and the small number of descriptors (five) selected to build the QSAR model.

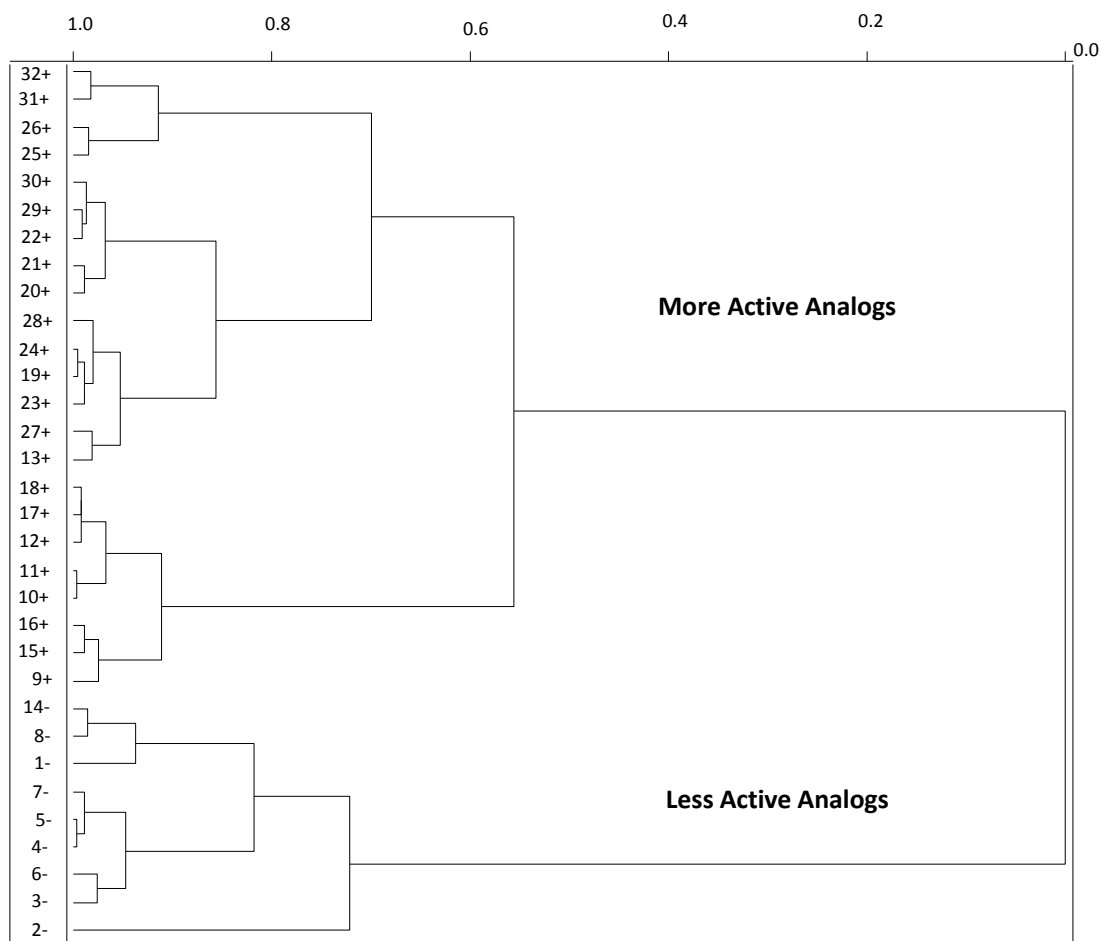


Fig. 5. HCA dendrogram for caffeine and derivatives with epithelial anticancer activity. Positive values indicate more potent analogs and negative values indicate less active compounds

3.4 Application of PLS and PCR Models to the Compounds of the Test Set

The compounds of the test set were modeled from the most stable structure of caffeine using GaussView 3.0 program. The optimization of the geometry of each compound was performed with DFT method and the basis set of separated valence 6-31G** by Gaussian 03 program. After obtain the most stable geometry of each compound was determined only selected descriptors in PCA and used in the building of the QSAR (PLS and PCR) models, namely area (A^2), distance radical 1 (dR1), distance radical 3 (dR3), radical partition coefficient 1 (logPR1) and radical partition coefficient 3 (logPR3), shown in Table 4.

The QSAR models (PLS and PCR) were used to predict the unknown epithelial anticancer activity

of fourteen new caffeine derivatives shown in Fig. 7, compounds 33-46. Table 5 shows the results of the ICT_{50} by PCR and PLS models. According to Table 5 the PLS model showed that eight compounds of the test set (37 and 40-46) are predicted to be more active, they had values of $ICT_{50} < 0.48$ mM. However, the PCR model only seven compounds of all test sets (33-36, 38, 39 and 42) were predicted as less active, which showed values of $ICT_{50} \geq 0.48$ mM, a total of 7 compounds proposed as more active of fourteen suggested compounds. However, compounds 37, 40, 41, 43, 44, 45 and 46 were the ones that had values of $ICT_{50} < 0.48$ mM in both models (PLS and PCR), suggesting that these new compounds in the two models are more potent than caffeine may be synthesized and tested for epithelial anticancer activity.

Table 3. Predicted PLS and PCR results and validation errors for ICT₅₀ (experimental)

| Compounds | Predicted | | Validation error | | Experimental |
|-----------|-----------|---------|------------------|---------|------------------------|
| | PLS | PCR | PLS | PCR | ICT ₅₀ (mM) |
| 1- | 0.5700 | 0.5962 | -0.0900 | -0.1162 | 0.4800 |
| 2- | 0.9583 | 0.8770 | 0.4283 | -0.347 | 0.5300 |
| 3- | 0.7892 | 0.8167 | 0.3908 | 0.3633 | 1.1800 |
| 4- | 0.5756 | 0.6193 | 0.1744 | 0.1307 | 0.7500 |
| 5- | 0.6628 | 0.7259 | 0.6072 | 0.5441 | 1.2700 |
| 6- | 0.7760 | 0.6858 | -0.286 | -0.1958 | 0.4900 |
| 7- | 0.6375 | 0.7065 | -0.1275 | -0.1965 | 0.5100 |
| 8- | 0.4830 | 0.4930 | 0.0770 | 0.0670 | 0.5600 |
| 9+ | 0.3180 | 0.3602 | 0.0620 | 0.0198 | 0.3800 |
| 10+ | 0.3404 | 0.3238 | -0.0904 | -0.0738 | 0.2500 |
| 11+ | 0.2935 | 0.3338 | 0.0665 | -0.0338 | 0.3000 |
| 12+ | 0.2281 | 0.1966 | -0.0981 | -0.0666 | 0.1300 |
| 13+ | 0.3027 | 0.3596 | -0.0027 | -0.0596 | 0.3000 |
| 14- | 0.5399 | 0.5213 | 0.1401 | 0.1587 | 0.6800 |
| 15+ | 0.3431 | 0.3125 | -0.0431 | -0.0431 | 0.3000 |
| 16+ | 0.2738 | 0.2596 | -0.0938 | -0.0796 | 0.1800 |
| 17+ | 0.2617 | 0.2343 | -0.2117 | -0.1843 | 0.0500 |
| 18+ | 0.2061 | 0.1810 | -0.1561 | -0.1310 | 0.0500 |
| 19+ | 0.1111 | 0.1457 | 0.0008 | -0.0257 | 0.1200 |
| 20+ | 0.1203 | 0.0752 | -0.0703 | -0.0252 | 0.0500 |
| 21+ | 0.2730 | 0.3236 | -0.1430 | -0.1936 | 0.1300 |
| 22+ | 0.0140 | -0.0189 | 0.1360 | 0.1689 | 0.1500 |
| 23+ | 0.1711 | 0.1238 | -0.1211 | -0.0738 | 0.0500 |
| 24+ | 0.0798 | 0.0648 | 0.0702 | 0.0852 | 0.1500 |
| 25+ | 0.0222 | -0.0021 | 0.0178 | 0.0421 | 0.0400 |
| 26+ | -0.0768 | -0.1208 | 0.1068 | 0.1508 | 0.0300 |
| 27+ | 0.1925 | 0.3046 | -0.1425 | -0.2546 | 0.0500 |
| 28+ | 0.0415 | 0.0265 | -0.0215 | 0.0065 | 0.0200 |
| 29+ | 0.0727 | 0.0100 | -0.0227 | 0.0400 | 0.0500 |
| 30+ | -0.0403 | -0.0562 | 0.0503 | 0.0662 | 0.0100 |
| 31+ | -0.0774 | -0.1130 | 0.1074 | 0.1430 | 0.0300 |
| 32+ | -0.1851 | -0.2226 | 0.2151 | 0.2526 | 0.0300 |

Table 4. Molecular properties selected by analysis of main components of test set with anticancer activity unknown

| Compounds | dR1 | dR3 | logPR1 | logPR3 | A ² |
|-----------|--------|--------|--------|--------|----------------|
| 33 | 1.0150 | 0.0000 | 0.0000 | 0.0000 | 279.2799 |
| 34 | 1.0140 | 2.1050 | 0.0000 | 1.0900 | 312.9500 |
| 35 | 1.0140 | 2.1050 | 0.0000 | 1.0900 | 338.0199 |
| 36 | 3.4370 | 2.0750 | 1.3000 | 1.0900 | 357.7099 |
| 37 | 3.4370 | 3.4340 | 1.3000 | 1.3000 | 381.5599 |
| 38 | 4.6770 | 0.0000 | 1.6900 | 0.0000 | 352.5299 |
| 39 | 1.0140 | 4.6730 | 0.0000 | 1.6900 | 365.8500 |
| 40 | 4.6770 | 2.1050 | 1.6900 | 1.0900 | 385.9500 |
| 41 | 2.1080 | 4.6730 | 1.0900 | 1.6900 | 408.0700 |
| 42 | 1.0140 | 4.6730 | 0.0000 | 1.6900 | 416.3999 |
| 43 | 1.0140 | 4.6730 | 0.0000 | 1.6900 | 443.4800 |
| 44 | 1.0140 | 5.9820 | 0.0000 | 2.0900 | 394.4899 |
| 45 | 7.2450 | 0.0000 | 2.4900 | 0.0000 | 415.9299 |
| 46 | 8.5440 | 0.0000 | 2.8800 | 0.0000 | 450.9700 |

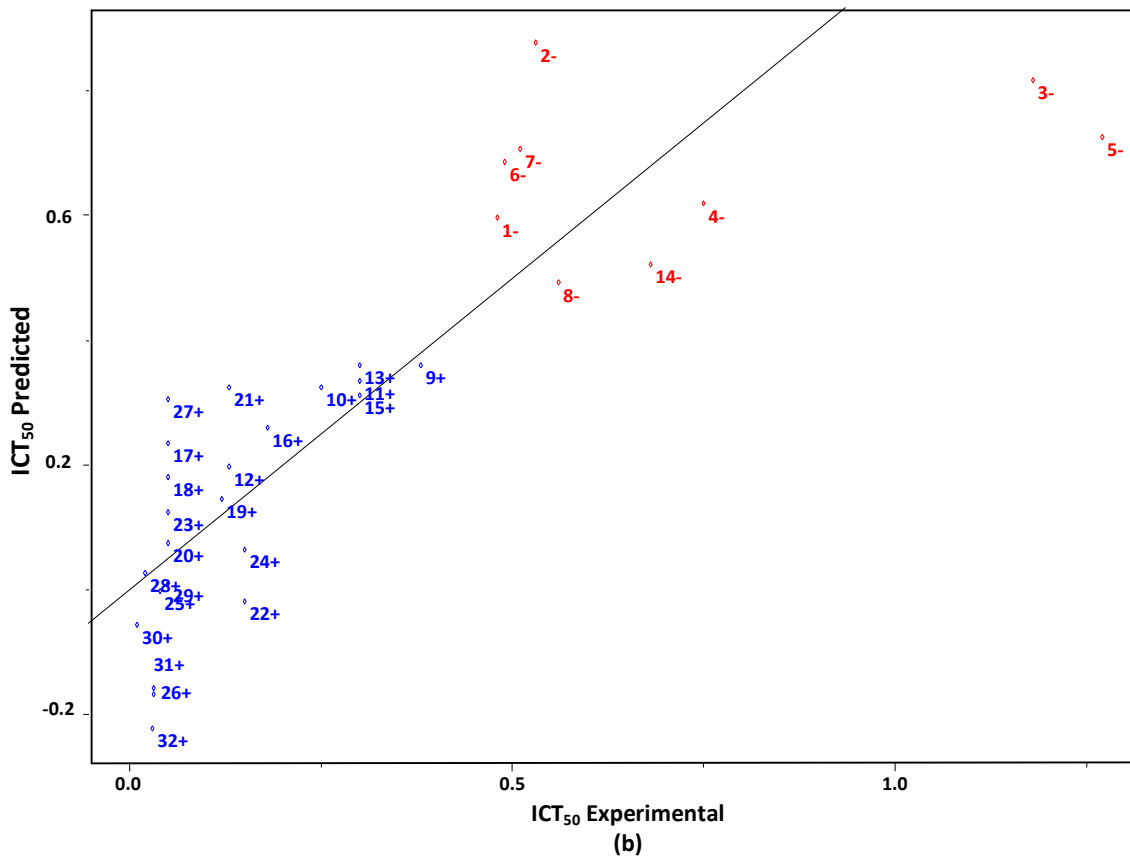
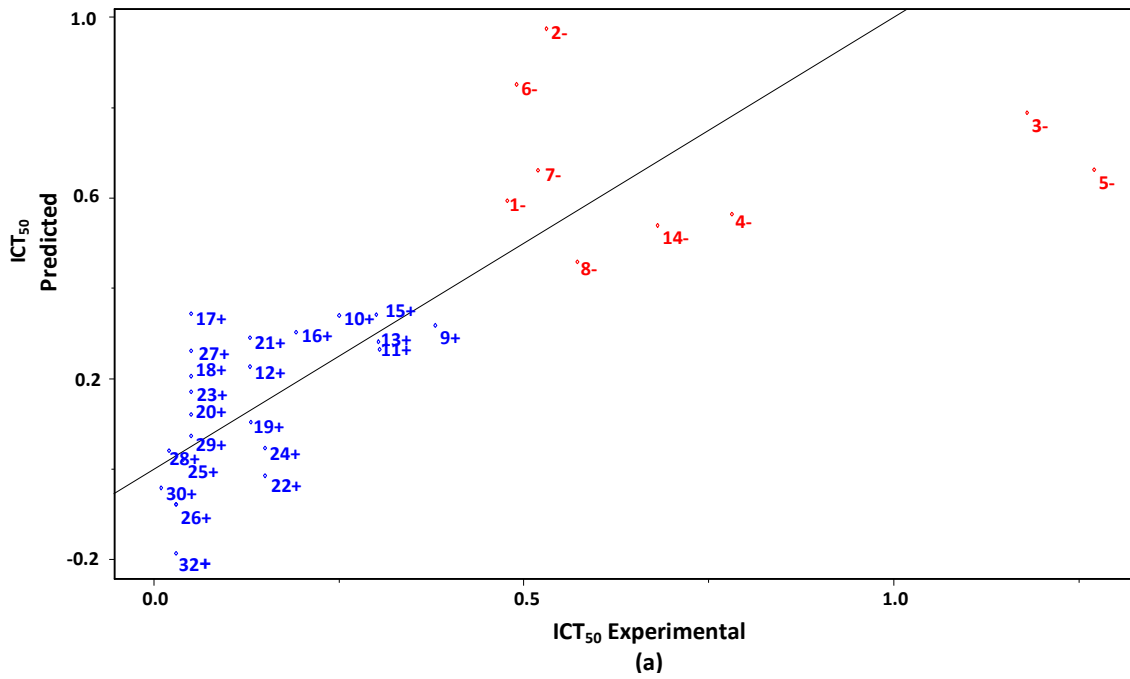


Fig. 6. Plot of experimental versus predicted values for ICT_{50} modeled by (a) PLS and (b) PCR

Table 5. Anticancer activity predicted (ICT₅₀) by PCR and PLS models for the test set compounds and residues of prediction between models

| Test set compounds | Predicted (ICT ₅₀) | | Residues of prediction (PLS-PCR) |
|--------------------|--------------------------------|--------|----------------------------------|
| | PLS | PCR | |
| 33 | 0.9261 | 0.9179 | 0.0082 |
| 34 | 0.7112 | 0.7261 | -0.0149 |
| 35 | 0.6840 | 0.6978 | -0.0138 |
| 36 | 0.5229 | 0.5156 | 0.0073 |
| 37 | 0.4318 | 0.4317 | 0.0001 |
| 38 | 0.6521 | 0.6127 | 0.0394 |
| 39 | 0.5101 | 0.5412 | -0.0311 |
| 40 | 0.4375 | 0.4212 | 0.0163 |
| 41 | 0.3674 | 0.3832 | -0.0158 |
| 42 | 0.4552 | 0.4840 | -0.0288 |
| 43 | 0.4258 | 0.4534 | -0.0276 |
| 44 | 0.3962 | 0.4370 | -0.0408 |
| 45 | 0.4725 | 0.4140 | 0.0585 |
| 46 | 0.3794 | 0.3112 | 0.0682 |

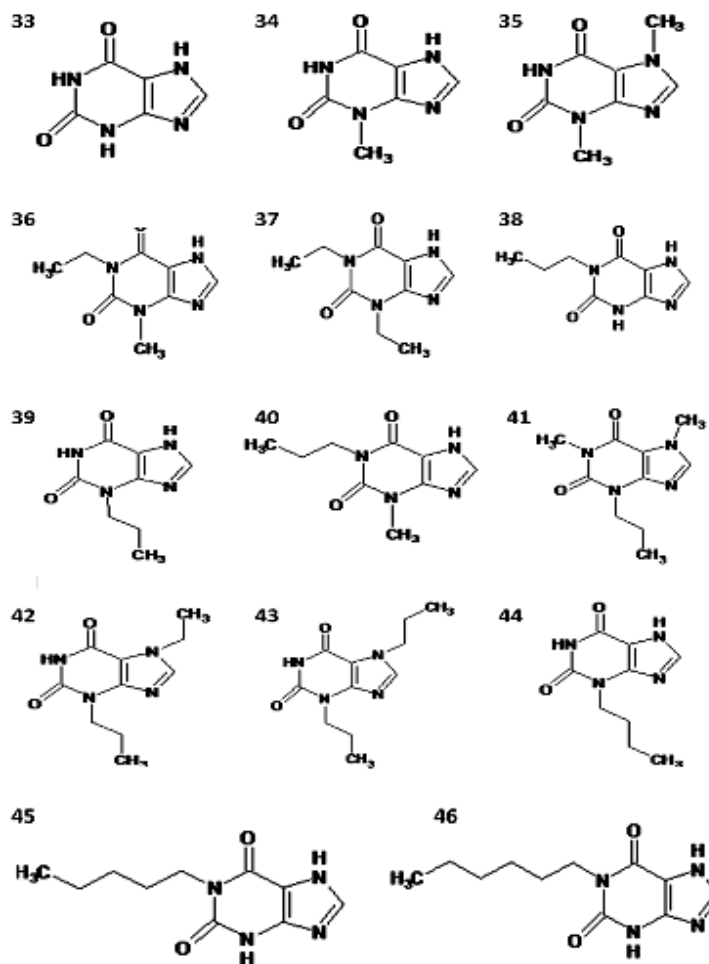


Fig. 7. Compounds of the test set caffeine derivatives with unknown anticancer activity against epithelial cancer

4. CONCLUSION

The DFT method and B3LYP/6-31G** basis set revealed themselves to be adequate to optimize the structures of caffeine and derivatives for subsequent study. PCA and HCA methods classified the compounds into groups according to their degree of epithelial anticancer activity. The descriptors dR1, dR3, logPR1, logPR3 and Δ^2 were responsible for distinguishing compounds with higher and lower activity. PLS and PCR models obtained showed good predictive ability. The test set showed for seven new caffeine compounds satisfactory results for epithelial anticancer activity. This strategy is fundamental for use in experimental syntheses and biological evaluation, and to understand the structural requirements for designing new ligands as anticancer agents. Consequently, further studies need be done to evaluate the different proposals as well as their actions, toxicity, and potential use for treatment of epithelial cancer.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

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COMPETING INTERESTS

The authors confirm that this article content has no conflicts of interest and part of the methodology of this article has been previously published in *Molecules* 2014, 19 (8), 10670-10697; doi:[10.3390/molecules190810670](https://doi.org/10.3390/molecules190810670).

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